# 1

### SURFACE MODIFICATION OF BIOPOLYMERS: AN OVERVIEW

Manju Kumari Thakur<sup>1</sup>, Ashvinder Kumar Rana<sup>2</sup>, Yang Liping<sup>3</sup>, Amar Singh Singha<sup>4</sup>, and Vijay Kumar Thakur<sup>5</sup>

<sup>1</sup>Division of Chemistry, Govt. Degree College Sarkaghat, Himachal Pradesh University, Shimla, Himachal Pradesh, India

<sup>2</sup>Department of Chemistry, Sri Sai University, Palampur; Himachal Pradesh, India

<sup>3</sup> Polymer Engineering and Catalysis, Institute of Chemical and Engineering Sciences, Singapore, Singapore

<sup>4</sup> Department of Chemistry, National Institute of Technology, Hamirpur, Himachal Pradesh, India <sup>5</sup> School of Mechanical and Materials Engineering, Washington State University, Pullman, WA, USA

#### 1.1 INTRODUCTION

Among various materials used in our everyday life, polymers play the most imperative role along with their use in a number of industries for versatile applications [1–3]. Polymers have been known to play a key role in the economy of the most of the countries of modern world since last century [4, 5]. Polymers have been frequently classified into natural and synthetic polymers [6, 7]. Although natural polymers were frequently used by the people of earlier civilization for a number of applications directly/indirectly, usage of synthetic polymers has dominated the modern world especially the last few decades [8–11]. Synthetic polymers have even replaced some of the commonly used metallic materials due to their enormous advantages such as light weight, chemical/water resistant, versatility, decent mechanical/thermal properties, and easy tailor ability [12–15]. Both natural and synthetic polymers can be easily distinguished depending upon their inherent properties and their structural property

*Surface Modification of Biopolymers*, First Edition. Edited by Vijay Kumar Thakur and Amar Singh Singha.

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relationship [16]. However, during the last few years, sustainable development through the use of environmentally friendly materials has become the hottest topic of conversation as well as research all around the globe [17, 18]. In this direction, the usage of materials that can be procured from our nature is rising rapidly for a number of applications [19, 20]. In fact the materials obtained from the nature are becoming a potentially viable alternative to a number of traditional synthetic materials that are generally synthesized from petroleum-based resources [21-23]. The effective utilization of the materials obtained from nature offers a number of challenges for their successful usage as well as offers new opportunities from the economic and environmental point of view [24, 25]. The past few decades has seen a pronounced advancement in the development of new eco-friendly materials that are procured from bio-based biopolymers for vast applications [26-28]. Different kinds of bio-based biopolymers represent a renewable feedstock of materials for different usage [29, 30]. The renewable feedstock of biopolymers extensively depends upon the availability of bio-based resources in different regions of the world, the new developments in the use of these materials, and the agricultural production as most of the biopolymers are directly/indirectly related to the field of agriculture [31, 32]. Applications of any biopolymer material in a particular application stresses on the specific physical, chemical, thermal, mechanical, economic, and degradation properties so as to offer significant advantages over their synthetic counterpart [33, 34]. In addition to these requirements, the easy availability of these biopolymeric materials is one of the most significant parameters in their commercialization as it is directly related to the final cost of the material in the market [35, 36]. Different kinds of biopolymer-based materials found in the nature can play one of the key roles in the modern industries to make the final product green [37, 38]. The use of biopolymer-based materials ranges from house hold applications to advanced applications in the defense [39, 40]. Different kinds of biopolymers depending upon their compositions can be used in a number of applications as follows: biomedical (e.g., stent, drug-delivery vehicles), food packaging, polymer composites for structural applications, as electrolyte for energy storage in super capacitor/battery, adhesives, cosmetic industries, and most frequently in textile industries [41, 42].

## **1.2 STRUCTURES OF SOME COMMERCIALLY IMPORTANT BIOPOLYMERS**

Among the various biopolymer materials, a few materials such as natural cellulosic fibers, starch, agar, chitosan, and poly(3-hydroxyalkanoates) (PHAs), are being used in a number of applications [24, 43–49]. In the following section, we briefly describe some of the commercially important biopolymers, as their detailed introduction along with their modification/applications has been given in the upcoming chapters.

#### 1.2.1 Natural Fibers

Among the various fibers available naturally/synthetically, natural cellulosic fibers are of much importance due to their intrinsic properties [48–50]. These fibers have been reported to be used by human beings for thousands of years ago starting from early civilization in the formation of bridges for on-foot passage as well as in naval



**FIGURE 1.1** Structure of lignocell ulosic natural fiber. Reproduced with permission from Ref. [51]. © 2013 Elsevier.



FIGURE 1.2 Structure of cellulose. Reproduced with permission from Refs. [48–50]. © Elsevier.

ships to biomedical in the present time [48–50]. Depending on their extraction as well as on the part of the plant from which they are taken, their properties vary considerably [50]. Figure 1.1 shows the schematic representation of natural fibers [51].

Natural cellulosic fibers primarily contain cellulose, hemicellulose, and lignin as their primary constituent and have been well researched as well as documented in the existing literature [48]. Figure 1.2 shows the structure of cellulose found in natural fibers. Cellulose (a nonbranched polysaccharide) is the prime constituent of all lignocellulosic natural fibers and has been found to exist in two crystalline forms, namely, cellulose I and II [48–51]. Cellulose is a linear condensation polysaccharide that comprises a D-anhydro glucopyranose units joined by  $\beta$ -1,4-glycosidic bonds. On the other hand, hemicelluloses are composed of a combination of 5- and 6-ring carbon ring sugars and have been found to remain associated with cellulose even after the removal of lignin [49, 50]. As opposed to the structure of cellulose, hemicelluloses exhibit a branched structure and consist of mixtures of polysaccharides with much lower molecular weight compared to cellulose [48–50].

Among different constituents of natural cellulosic fibers, lignin is one of the highly branched components. It is a complex chemical compound present in huge quantities in the cell walls of plants. It is the main binding agent for components of the plants and serves as a matrix to the embedded cellulose fibers along with hemicellulose. The structure of lignin is highly branched that consists of phenyl propane units. These units are organized in a complex three-dimensional structure linked together through numerous types of carbon–carbon and ether bonds.

#### 1.2.2 Chitosan

Chitosan is another most significant biopolymer that is derived from chitin (produced by many living organisms) [1, 3, 52]. Chitin is the second most abundant natural polymer available on earth after cellulose and is found in a number of organisms from crustaceans such as lobsters, crabs, shrimp, and prawns along with insects to some types of fungi [31, 34, 52]. Chitin is a nitrogen-rich polysaccharide and a high-molecular-weight linear polymer composed of *N*-acetyl-D-glucosamine (*N*-acetyl-2-amino-2-deoxy-D-glucopyranose) units linked by  $\beta$ -D-(1 $\rightarrow$ 4) bonds. Figure 1.3 shows the comparative chemical structure of chitin, chitosan, and cellulose [49, 52].



**FIGURE 1.3** Structure of chitosan, chitin, and cellulose. Reproduced with permission from Ref. [52]. © 2013 Elsevier.



**FIGURE 1.4** Deacetylation of chitin to chitosan. Reproduced with permission from Ref. [52]. © 2013 Elsevier.

Chitosan is most frequently produced from chitin by deacetylation process [31, 34, 49, 52]. Figure 1.4 shows the scheme for the extraction of chitosan from chitin.

Chitosan has been found to exhibit a basic character and is one of the highly basic polysaccharides compared to other natural polysaccharides that are acidic such as cellulose, agar, pectin, dextrin, and agarose [24, 29–53]. The degree of deacetylation and the charge neutralization of  $-NH_2$  groups along with ionic strength have been found to control the intrinsic  $pK_a$  value of chitosan. Figure 1.5 shows the schematic illustration of the versatility of chitosan.

Along with physical modification, chitosan is most commonly modified by a number of chemical techniques including traditional chemical modification techniques such as photochemical, enzymatic, radiation, and plasma-induced graft copolymerization. Figure 1.6 summarizes some of the commercially used modification techniques for surface modification of chitosans.

#### 1.2.3 Agar

Agar has been commonly recognized as a hydrophilic colloid that is most frequently extracted from certain marine algae and is accumulated in the cell walls [2]. Due to its significant biological activities such as antioxidative, anticancer, anticoagulant, antiviral,



**FIGURE 1.5** Schematic illustration of chitosan's versatility. At high pH (above 6.5), chitosan's amine groups are deprotonated and reactive. At low pH (<6.7), chitosan's amines are protonated, confirming the polycationic behavior of chitosan. Reproduced with permission from Ref. [52]. © 2013 Elsevier.



**FIGURE 1.6** Chemical modification of chitosan for different applications: (a) methylation, (b) thiolation, (c) azylation, (d) copolymerization, and (e) N-succinylation. Reproduced with permission from Ref. [52]. © 2013 Elsevier.



**FIGURE 1.7** Structural motifs of agar polysaccharides showing carbon numbering  $(C_1 - C_6)$ . Reproduced with permission from Ref. [53]. © 2013 Elsevier.

and immunomodulating activities, it is extensively used as a gelling/stabilizing agent in a number of industries starting from food to pharmaceutical. Figure 1.7 shows the structural motifs of agar polysaccharides showing carbon numbering  $(C_1-C_6)$ .

Agar is represented by the structural formula  $(C_{12}H_{18}O_9)_n$  and is built on a disaccharide-repeating unit of 3-linked β-D-galactose (G) and 4-linked 3,6-anhydro-α-Lgalactose (AG) residues. Depending on the source of the agar, there is possible occurrence of different substituents such as sulfate, methoxyl, and/or pyruvate at various positions in the polysaccharide chain. Agar is extracted from macroalgae by a number of conventional techniques, and recently great efforts are being made to extract it rapidly using a microwave-assisted methodology [32, 43, 44, 53]. Depending on the targeted applications, agar is surface modified accordingly [43, 44]. For example, agar is extracted after the alkali treatment to be used as catalysts supports and templates for the preparation of metal oxides using the same methodology and has been characterized by a number of techniques such as X-ray photoelectron spectroscopy (XPS), scanning electron microscopy (SEM), nuclear magnetic resonance spectroscopy (CP-MAS <sup>13</sup>C NMR and diffuse reflectance infrared Fourier transform spectroscopy (DRIFTS)) [53]. Figure 1.8 shows the SEM images of the alkali treated and untreated agar. These images clearly illustrate the structural changes taking place as a result of alkali treatment.



**FIGURE 1.8** SEM micrographs of nontreated (a) and alkali-treated (b) mesoporous agar materials. Reproduced with permission from Ref. [53]. © 2013 Elsevier.

The results obtained from the SEM images were further confirmed by DRIFTS and NMR. Figure 1.9 shows the presence of different functional groups on both the treated and untreated agar.

Figure 1.10 also confirms the agarose structure of the extracted material after the alkali treatment. The structures of both the raw and alkali-treated agar show significant changes in their peaks.

#### 1.4 POLY(3-HYDROXYALKANOATES)

PHAs are one of the unique families of polymers that are gaining significant attention during the past few years [49, 54, 55]. PHAs are linear, biodegradable polyesters that are synthesized by a wide variety of bacteria through the fermentation of sugars, lipids, among others [49, 54, 55]. Due to their biodegradable, renewable, and biocompatible



**FIGURE 1.9** Attenuated total reflectance infrared (FTIR-ATR) spectra of nontreated (a) and alkali-treated (b) agar extracted from *Gracilaria gracilis*. Reproduced with permission from Ref. [53]. © 2013 Elsevier.

nature, PHAs are emerging as one of the most promising materials for biomedical applications [49, 54, 55]. The structure and different applications of PHAs have been discussed in some of the recent reviews. Similar to other previously discussed polymers, PHAs also suffer from few drawbacks. Figure 1.11 shows the structural representation for different kinds of PHAs [54]. Some of the disadvantages of the PHAs include the brittleness and poor mechanical properties that restrict its successful applications [49, 54, 55]. Therefore to get the desired properties, it is most of the time modified with suitable materials/techniques.



**FIGURE 1.10** CP-MAS <sup>13</sup>C NMR spectra of native agar (a) extracted at 100°C and alkalitreated agar (b) extracted at 140°C from *Gracilaria*. Top structure depicts the various carbons  $(C_1-C_6 \text{ from G and AG})$  associated with the different NMR peaks. Reproduced with permission from Ref. [53]. © 2013 Elsevier.

The effect of microstructure and composition of PHAs, along with the influence of compositional distribution and blending on mechanical properties, has been reported in detail by a number of researchers [49, 54, 55]. Figure 1.12 schematically shows the biochemistry and crystallinity of the PHAs, as these control most of the properties of the resulting material [55].





**FIGURE 1.11** Chemical structure of PHAs. Reproduced with permission from Ref. [54]. © 2013 Elsevier.



**FIGURE 1.12** Overview of PHA synthesis: schematic depiction of (a) chain polymerization catalyzed by enzymes, (b) a PHA granule with granule-associated proteins, (c) different forms of the PHB polymer chain, and (d) semicrystalline polymer structure. (e) AFM image of PHBV film; (f) final plastic products. Reproduced with permission from Ref. [55]. © 2013 Elsevier. (*See insert for color representation of the figure.*)

A number of mechanisms have been proposed for the synthesis of PHAs from different precursors (Figure 1.13; [55]).

As it is evident from the existing literature that PHAs alone cannot satisfy the strict requirements for certain applications, most of the time it is blended with other materials. It has been reported that such blends exhibit a number of properties ranging from complete cocrystallization through partial segregation. Figure 1.14 shows the variation in phase structure of some of the blends.

(a)



**FIGURE 1.13** Proposed polymerization mechanism for the synthesis of PHA. Reproduced with permission from Ref. [55]. © 2013 Elsevier.



**FIGURE 1.14** Variation of phase structures in PHB/PHB-HV blends. Reproduced with permission from Ref. [55]. © 2013 Elsevier.

Although these polymers have some disadvantages, they are promising candidates as novel material for petroleum-based materials with the advantages of being biodegradable.

#### 1.5 STARCH

Among various polysaccharides, starch is an industrially important biopolymer frequently used in food industry as well as in other industries. Depending on the targeted applications, starch has been used as a colloid stabilizer, thickener, and bulking/ gelling agent [56]. Due to its enormous advantages, nearly 60 million tons of production has been reported all over the world [56]. Figure 1.15 shows the common structure sketches of starch granules. Starch has been found to be composed of different kinds of glucoses (primarily amylose (AM) and amylopectin (AP)) and occurs in the form of granules.

Starch is classified into normal, wx, and high AM depending upon the amount/ ratio of amylose/amylopectin. Although starch is available in large quantity and offers numerous advantages, it has also some shortcomings similar to other biopolymers and has limited applications in its native form. A number of surface modification techniques are being used to alter the surface properties of starch and has been described in detail in the extensive reviews by Pei-Ling as well as in Chapter 7 in this book. Among these, one of the most promising is high hydrostatic pressure (HHP),



Corn starch granule is polyhedron with many flat surfaces and edge angles



which is a well-established nonthermal processing technology. Figure 1.16 shows the A- and B-type polymorphic structure of amylase as a result of HHP processing [56].

As a result of HHP processing, the structure of starch has also been found to be affected. Figure 1.17 shows the morphological images of the starch processed under high pressure. It is obvious from the micrographs that the outer surface of the starch granule is quite resistant to the HHP, and at the same time, the inner part is filled with structure having the shape of gel.

From the above discussion, it is quite clear that each biopolymer has certain limitation that restricts its use for a number of applications. The main obstacles include the lack of desired physicochemical/mechanical and biological properties. The only way to effectively utilize them is to alter their properties by surface modification



**FIGURE 1.16** A- and B-type polymorphs of amylase. Reprinted with permission from Ref. [56].  $\bigcirc$  2010 Wiley-VCH.



**FIGURE 1.17** SEM microstructure of potato starch with details of outer and inner part of starch structure: native (a); treated with high pressure at 600 MPa 3 min (b–d). Reprinted with permission from Ref. [56]. © 2010 Wiley-VCH.

techniques. By utilizing different surface modifications, most of the times, the specific application properties of different biopolymers can be easily obtained. For a particular application, the analysis of the structure–property relationship of a biopolymer under investigation is of utmost importance. At present, a number of biopolymers are being tested for their commercial applications and some of the thrust areas include biomedical, packaging, food production, and automotive. Some biopolymers can directly replace synthetically derived materials in traditional applications, whereas others possess unique properties that could open up a good range of new commercial opportunities. In this book, best efforts have been made to incorporate sufficient information on different surface modification techniques to alter their specific properties for targeted applications.

#### REFERENCES

- Depan D, Shah JS, Misra RDK. Degradation mechanism and increased stability of chitosan-based hybrid scaffolds cross-linked with nanostructured carbon: process-structurefunctional property relationship. *Polym Degrad Stab* 2013;98 (11):2331–2339.
- [2] Thakur VK, Ding G, Ma J, Lee PS, Lu X. Hybrid materials and polymer electrolytes for electrochromic device applications. *Adv Mater* 2012;24:4071–4096.
- [3] Depan D, Kumar AP, Singh RP. Preparation and characterization of novel hybrid of chitosan-g-lactic acid and montmorillonite. J Biomed Mater Res A 2006;78:372.
- [4] Thakur VK, Tan EJ, Lin MF, Lee PS. Poly (vinylidene fluoride)—graft—poly(2-Hydroxyethyl methacrylate): novel material for high energy density capacitors. J Mater Chem 2011;21:3751–3759.
- [5] Thakur VK, Thakur MK, Gupta RK. Synthesis of lignocellulosic polymer with improved chemical resistance through free radical polymerization. *Int J Biol Macromol* 2013;61:121–126.
- [6] Thakur VK, Singha AS, Thakur MK. Graft copolymerization of methyl acrylate onto cellulosic biofibers: synthesis, characterization and applications. J Polym Environ 2012;20 (1):164–174.
- [7] Thakur VK, Tan EJ, Lin MF, Lee PS. Polystyrene grafted polyvinylidenefluoride copolymers with high capacitive performance. *Polym Chem* 2011;2:2000–2009.
- [8] Thakur VK, Thakur MK, Gupta RK. Rapid synthesis of graft copolymers from natural cellulose fibers. *Carbohydr Polym* 2013;98:820–828.
- [9] Thakur VK, Thakur MK, Gupta RK. Graft copolymers from cellulose: synthesis, characterization and evaluation. *Carbohydr Polym* 2013;97:18–25.
- [10] Thakur VK, Singha AS. Physico-chemical and mechanical behavior of cellulosic pine needles based biocomposites. *Int J Polym Anal Charact* 2011;16:390–398.
- [11] Khoathane MC, Sadiku ER, Wambua PM. Effect of water glass treatment on the mechanical and thermooxidative properties of kenaf and sisal fibres. J Reinf Plast Compos 2013;31:1261–1269.
- [12] Dhakal HN, Zhang ZY, Bennett N. Influence of fibre treatment and glass fibre hybridisation on thermal degradation and surface energy characteristics of hemp/unsaturated polyester composites. *Compos Part B* 2012;43:2757–2761.

- [13] Thakur VK, Yan J, Lin MF, Zhi C, Golberg D, Bando Y, Sim R, Lee PS. Novel polymer nanocomposites from bioinspired green aqueous functionalization of BNNTs. *Polym Chem* 2012;3:962–969.
- [14] Thakur VK, Lin MF, Tan EJ, Lee PS. Green aqueous modification of fluoropolymers for energy storage applications. J Mater Chem 2012;22:5951–5959.
- [15] Singha AS, Thakur VK, Mishra BN. Study of Grewia optiva fiber reinforced urea-formaldehyde composites. J Polym Mater 2009;26 (1):81–90.
- [16] Thakur VK, Thakur MK, Gupta RK. Development of functionalized cellulosic biopolymers by graft copolymerization. *Int J Biol Macromol* 2013;62:44–51.
- [17] Depan D, Kumar AP, Singh RP. Cell proliferation and controlled drug release studies of nanohybrids based on chitosan-g-lactic acid and montmorillonite. *Acta Biomater* 2009; 5:93–100.
- [18] Sousa Neto VO, Carvalho TV, Honorato SB, Gomes CL, Barros FCF, Araújo-Silva MA, Freire PTC, Nascimento RF. Coconut bagasse treated by thiourea/ammonia solution for cadmium removal: kinetics and adsorption equilibrium. *BioResources* 2012;7: 1504–1524.
- [19] Mishra S, Rani GU, Sen G. Microwave initiated synthesis and application of poly acrylic acid grafted carboxymethylcellulose. *Carbohydr Polym* 2012;87:2255–2262.
- [20] Aldana AA, Toselli R, Strumia MC, Martinelli M. Chitosan films modified selectively on one side with dendritic molecules. J Mater Chem 2012;22:22670–22677.
- [21] Sousa FW, Moreira SA, Oliveira AG, Cavalcante RM, Rosa MF, Nascimento RF. The use of green coconut shells as absorbents in the toxic metals. *Quim Nova* 2007;30: 1153–1157.
- [22] Thakur VK, Singha AS. Rapid synthesis, characterization, and physicochemical analysis of biopolymer-based graft copolymers. *Int J Polym Anal Charact* 2011;16:153–164.
- [23] Depan D, Kumar B, Singh RP. Preparation and characterization of novel hybrid of chitosan-g-PDMS and sodium montmorillonite. J Biomed Mater Res B Appl Biomater 2008;84:184.
- [24] Thakur VK, Singha AS, Thakur MK. In-air graft copolymerization of ethyl acrylate onto natural cellulosic polymers. *Int J Polym Anal Charact* 2012;17:48–60.
- [25] Grande D, Renard E, Babinot J, Ramier J, Langlois V. Harnessing biopolyesters in the design of functional and nanostructured architectures. In: Khemani K, Scholz C, editors. Degradable Polymers and Materials : Principles and Practice. 2nd ed. Washington: American Chemical Society; 2012. ACS symposium series; p 187–199.
- [26] Sousa Neto VO, Oliveira AG, Teixeira RNP, Silva MAA, Freire PTC, Keukeleire DD, Nascimento RF. Use of coconut bagasse as alternative adsorbent for separation of copper (II) ions from aqueous solutions: isotherms, kinetics, and thermodynamic studies. *BioResources* 2011;6:3376–3395.
- [27] Lao HK, Renard E, Linossier I, Langlois V, Vallee-Rehel K. Modification of poly(3hydroxybutyrate-co-3-hydroxyvalerate) film by chemical graft copolymerization. *Biomacromolecules* 2007;8:416–423.
- [28] Avila A, Bierbrauer K, Pucci G, Lopez-Gonzalez M, Strumia M. Study of optimization of the synthesis and properties of biocomposite films based on grafted chitosan. J Food Eng 2012;109:752–761.
- [29] Versace D-L, Ramier J, Babinot J, Lemechko P, Soppera O, Lalevee J, Albanese P, Renard E, Langlois V. Photoinduced modification of the natural biopolymer poly

(3-hydroxybutyrate-co-3-hydroxyvalerate) microfibrous surface with anthraquinonederived dextran for biological applications. *J Mater Chem B* 2013;1:4834–4844.

- [30] Capra RH, Baruzzi AM, Quinzani LM, Strumia MC. Rheological, dielectric and diffusion analysis of mucin/carbopol matrices used in amperometric biosensors. *Sens Actuators B* 2007;124:466–476.
- [31] Depan D, Venkata Surya PKC, Misra RDK. Organic/inorganic hybrid network structure nanocomposite scaffolds based on grafted chitosan for tissue engineering. *Acta Biomater* 2011;7:2163.
- [32] Rani GU, Mishra S, Sen G, Jha U. Polyacrylamide grafted agar: synthesis and applications of conventional and microwave assisted technique. *Carbohydr Polym* 2012;90: 784–791.
- [33] Lao HK, Renard E, Langlois V, Vallee-Rehel K, Linossier I. Surface functionalization of PHBV by HEMA grafting via UV treatment: comparison with thermal free radical polymerization. J Appl Polym Sci 2010;116:288–297.
- [34] Depan D, Misra RDK. The interplay between nanostructured carbon-grafted chitosan scaffolds and protein adsorption on the cellular response of osteoblasts: structurefunction property relationship. *Acta Biomater* 2013;9:6084.
- [35] Shibi IG, Anirudhan TS. Synthesis, characterization, and application as a mercury (II) sorbent of banana stalk (Musa paradisiaca)-polyacrylamide grafted copolymer bearing carboxyl groups. *Ind Eng Chem Res* 2002;41:5341–5352.
- [36] Mishra S, Sen G. Microwave initiated synthesis of polymethylmethacrylate grafted guar (GG-g-PMMA), characterizations and application. *Int J Biol Macromol* 2011;48: 688–694.
- [37] Capra RH, Strumia M, Vadgama PM, Baruzzi AM. Mucin/carbopol matrix to immobilize oxalate oxidase in a urine oxalate amperometric biosensor. *Anal Chim Acta* 2005;530:49–54.
- [38] Renard E, Walls M, Guerin P, Langlois V. Hydrolytic degradation of blends of polyhydroxyalkanoates and functionalized polyhydroxyalkanoates. *Polym Degrad Stab* 2004;85:779–787.
- [39] Bear MM, Mallarde D, Langlois V, Randriamahefa S, Bouvet O, Guerin P. Natural and artificial functionalized biopolyesters. II. Medium-chain length polyhydroxyoctanoates from Pseudomonas strains. *J Polym Environ* 1999;7:179–184.
- [40] Sen G, Mishra S, Jha U, Pal S. Microwave initiated synthesis of polyacrylamide grafted guar gum (GG-g-PAM)–characterizations and application as matrix for controlled release of 5-amino salicylic acid. *Int J Biol Macromol* 2010;47:164–170.
- [41] Anirudhan TS, Sandeep S. Synthesis and characterization of molecularly imprinted polymer of N-maleoylchitosan-grafted-2-acrylamido-2-methylpropanesulfonic acid and its controlled delivery and recognition of bovine serum albumin. *Polym Chem* 2011;2: 2052–2061.
- [42] Sen G, Mishra S, Rani GU, Rani P, Prasad R. Microwave initiated synthesis of polyacrylamide grafted Psyllium and its application as a flocculant. *Int J Biol Macromol* 2012;50:369–375.
- [43] Rani GU, Mishra S, Pathak G, Jha U, Sen G. Synthesis and applications of poly(2hydroxyethylmethacrylate) grafted agar: a microwave based approach. *Int J Biol Macromol* 2013;61:276–284.

- [44] Sen G, Rani GU, Mishra S. Microwave assisted synthesis of poly(2-hydroxyethylmethacrylate) grafted agar (Ag-g-P(HEMA)) and its application as flocculant for wastewater treatment. *Front Chem Sci Eng* 2013;7:312–321.
- [45] Sousa FW, Oliveira AG, Ribeiro JP, Keukeleire D, Sousa AF, Nascimento RF. Single and multi-elementary isotherms of toxic metals in aqueous solution using treated coconut shell powder. *Desalin Water Treat* 2011;36:289–296.
- [46] Sousa FW, Sousa MJ, Oliveira IRN, Oliveira AG, Cavalcante RM, Fechine PBA, Sousa Neto VO, Keukeleire D, Nascimento RF. Evaluation of a low-cost adsorbent for removal of toxic metal ions from wastewater of an electroplating factory. *J Environ Manage* 2009;90:3340–3344.
- [47] Guérin P, Renard E, Langlois V. Degradation of natural and artificial poly[(R)-3hydroxyalkanoate]s: from biodegradation to hydrolysis. In: Chen GQ, editor. Plastics From Bacteria—Natural Functions and Applications. Berlin: Springer; 2010. p 283–321.
- [48] Thakur VK. Green Composites From Natural Resources. Boca Raton: CRC Press/ Taylor & Francis; 2013. p 1–419.
- [49] Singha AS, Thakur VK. Green Polymer Materials. Houston: Studium Press LLC; 2011. p 1–316.
- [50] Thakur VK, Singha AS. Biomass-Based Biocomposites. Shrewsbury: Smithers Rapra; 2013. p 1–386.
- [51] Rong ZM, Zhang QM, Liu Y, Gui Cheng Yang CG, Zeng HM. The effect of fiber treatment on the mechanical properties of unidirectional sisal-reinforced epoxy composites. *Compos Sci Technol* 2001;61:1437–1447.
- [52] Shukla SK, Mishra AK, Arotiba OA, Mamba BB. Chitosan-based nanomaterials: a stateof-the-art review. *Int J Biol Macromol* 2013;59:46–58.
- [53] Francavill M, Pineda A, Lin SK, Franchia M, Trottaa P, Romerob AA, Luque R. Natural porous agar materials from macroalgae. *Carbohydr Polym* 2013;92:1555–1560.
- [54] Hazer DB, Kılıçay E, Hazer B. Poly (3-hydroxyalkanoates): diversification and biomedical applications: a state of the art review. *Mater Sci Eng C* 2012;32:637–647.
- [55] Laycock B, Halley P, Pratt S, Werkerc A, Lanta P. The chemomechanical properties of microbial polyhydroxyalkanoates. *Prog Polym Sci* 2013;38:536–583.
- [56] Pei-Ling L, Xiao-Song H, Shen Q. Effect of high hydrostatic pressure on starches: a review. *Starch-Starke* 2010;62:615–628.