

Section I

INTRODUCTION

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Chapter 1

PHOSPHORUS: BACK TO THE ROOTS

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Abstract: Phosphorus (P) is a pivotal nutrient for all life on Earth. It is poorly mobile in soil and inorganic P concentrations in the soil solution are <0.6 to 11 μM . Organic P concentrations in the soil solution tend to be higher, but organic P needs to be hydrolysed before it can be taken up by plant roots. Such hydrolysis involves phosphatases that are either released from the roots or derived from microorganisms in the rhizosphere. A large fraction of soil P is sorbed onto soil particles, and hence is unavailable to most plants. Roots that release large amounts of P-solubilising carboxylates can access some of this sorbed P. Rates of P uptake from the soil solution are determined predominantly by the movement of P in soil. Root traits that enhance P movement in soil increase P acquisition; however, the kinetic properties of P transporters that take up this P have little effect on net P uptake. The downregulation of genes encoding these transporters is important to avoid P toxicity at a high P supply. Species or genotypes that lack the capacity to downregulate their P-uptake capacity typically show P-toxicity symptoms at a high P supply. Mycorrhizal symbionts increase the soil volume that is available for P acquisition. Attempts to select or engineer genotypes with greater P-uptake capacities should consider both root and soil characteristics, including soil microorganisms such as mycorrhizal fungi.

Keywords: Peak phosphorus, phosphorus mobility in soil, phosphorus toxicity, rhizosphere, soil phosphorus pools, sorption

1.1 Introduction

Phosphorus (P) is one of the major elements required by all living cells to grow and develop. Phosphorus does not occur naturally as a free element,

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because it is too reactive, combining rapidly with other elements such as oxygen or hydrogen. When P is oxidised to the fullest extent possible, the product is orthophosphate (PO_4^{3-} ; Pi), in which four oxygen atoms have bonded with a single P atom. At neutral pH, the Pi anion is present as a mixture of HPO_4^{2-} and H_2PO_4^- ; it is predominantly as H_2PO_4^- that P is transported into plant cells. The assimilated P is intimately involved with cellular bioenergetics and metabolic regulation, and is also an important structural component of essential biomolecules such as DNA, RNA, phospholipids (Chapter 9), ATP, and sugar-phosphates. Unlike in some bacterial cells, however, Pi cannot be reduced within plant cells to lower oxidation states. Rather, assimilated Pi is either sequestered in the cell vacuole or rapidly incorporated into organic form (e.g. initially as ATP) via photophosphorylation or oxidative phosphorylation.

Phosphorus plays a central role in virtually all major metabolic processes in plants, particularly photosynthesis and respiration, but it is also one of the least available macronutrients in many terrestrial and aquatic ecosystems. The massive use of P-containing fertilizers in agriculture, currently estimated to be in excess of 160 million metric tonnes (Mt) of rock phosphate (which equates to approximately 21 million Mt of P) per year worldwide (Fixen & Johnston, 2012), demonstrates how the plant-available P level of many soils is suboptimal for crop growth (Johnston *et al.*, 2014). The use of P-fertilizers can also be quite inefficient, with less than 20% of P applied to land that was recently converted to be used for crops or pastures typically being absorbed by plants during their first growing seasons (Simpson *et al.*, 2011). The judicious use of P fertilizer on established fields, however, allows 80–90% of applied P to be used by the crop and removed at harvest (Johnston *et al.*, 2014). The remaining P is sorbed onto soil particles or erodes and leaches (Andersson *et al.*, 2013; Smith & Schindler, 2009). Agricultural P runoff is a primary factor in the eutrophication of lakes and estuaries, and has also resulted in blooms of toxic cyanobacteria. With the world's population continuing its rapid increase, humankind faces a daunting challenge to produce sufficient food crops in the face of dwindling supplies of P-fertilizers. Thus, research on plant metabolic adaptations to suboptimal soil P availability, which is the focus of the present volume, is of significant practical importance. This will help to facilitate the development of effective tools and strategies for the rational application of biotechnology to reduce agriculture's current heavy reliance on expensive, polluting, and unsustainable P-fertilizers.

1.2 Phosphorus or phosphorous?

Phosphorus (P) must be the most frequently misspelled nutrient on our planet. In mainstream journals dealing with plant or soil science, the element is often misspelled as *phosphorous*. Spell-checkers do not pick up this mistake, however, because *phosphorous* is an existing word. It is not an

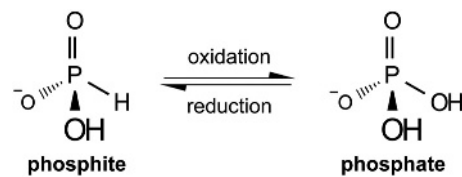


Figure 1.1 Phosphite and phosphate. Phosphite is less-oxidised than phosphate (Pi), and is not a direct source of P for plant nutrition. In soil, phosphite can be oxidised by microbes to Pi, which then makes it available for uptake by plant roots.

alternative spelling for *phosphorus*, however, and this is not a matter of British or American English. *Phosphorous acid* ($\text{HPO}(\text{OH})_2$) is a reduced form of Pi in which an oxygen bonded to the P atom is replaced by a hydrogen (Figure 1.1). Phosphite (H_2PO_3^- ; also known as phosphonate) is an alkali metal salt of phosphorous acid that represents an important but highly controversial agronomic commodity that is being widely marketed either as an agricultural fungicide or as a superior source of P for crop nutrition (McDonald *et al.*, 2001; Thao & Yamakawa, 2009). Published research conclusively indicates that phosphite functions as an effective control agent for a number of crop diseases caused by various species of pathogenic oomycetes belonging to the genus *Phytophthora* (e.g. Dunne *et al.*, 2010; McDonald *et al.*, 2001; Ratjen & Gerendás, 2009; Thao & Yamakawa, 2009). However, evidence that phosphite can be used directly by plants as a source of nutritional P is lacking. Phosphite can have direct effects on plants, because phosphite concentrations comparable to those required to control plant infection by pathogenic *Phytophthora* species are extremely phytotoxic to P-deprived plants; it is much less phytotoxic to P-fertilised plants (Carswell *et al.*, 1996; McDonald *et al.*, 2001; Ratjen & Gerendás, 2009; Thao & Yamakawa, 2009). This is because phosphite treatment effectively blocks the signal-transduction pathway by which plants (and yeast) perceive and respond to P-deprivation at the molecular level (Chapter 2). Thus, phosphite intensifies the deleterious effects of P-deficiency by 'tricking' P-deprived plant cells into sensing that they are P-sufficient when, in fact, their cellular P concentration is very low. Names are important, and so is characterisation of the mechanisms by which growth-enhancing substances actually work. Calling phosphite an agricultural fungicide in order to register it involves abiding by time-consuming and costly regulatory protocols. Calling phosphite a 'plant P fertilizer' can avoid the substantial expenses and tests associated with registering it as a fungicide. Whilst a number of agrochemical companies continue to market phosphite as a 'superior P fertilizer', a compound that suppresses P-starvation responses deserves the term of an 'anti-fertilizer' and should not be allowed to be advertised as an alternative to Pi (McDonald *et al.*, 2001; Ratjen & Gerendás, 2009; Thao & Yamakawa, 2009). However, as discussed in Chapter 2, the phosphite anion represents a useful tool to help dissect the signal-transduction

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pathways by which plants respond to nutritional P deprivation at the molecular level.

1.3 Phosphorus on a geological time scale

The cycling of P in the global environment is an important biogeochemical process. Phosphorus is present in only minute quantities in the Earth's crust (0.9 mg g^{-1} dry soil) (Filippelli, 2008). This is less than half of what is found in Martian rocks (Greenwood & Blake, 2006). Both, on Earth and on Mars, P is derived from the weathering of igneous rocks. It has been suggested that the greatly increased oxygen concentration of the Earth's atmosphere that occurred between 2.3 and 2.4 billion years ago (the so-called 'Great Oxidation Event') (Canfield *et al.*, 2013), may have been the cause of enhanced oxidative weathering on land. This oxidative weathering liberated more P to the oceans and stimulated primary production and organic carbon burial (Bekker & Holland, 2012). Because of the relatively small quantities in the Earth's crust, it took about three billion years before enough P was weathered from igneous rock in which it was entrapped for the seas to become saturated (Griffith *et al.*, 1977). This led to the formation of phosphate rock reserves that are now mined to produce P fertilizers (Cooper *et al.*, 2011).

The roots of early terrestrial plants, which occurred on soils with relatively low availability of P, evolved symbioses with mycorrhizal fungi in order to acquire sufficient soil P more than 400 million years ago (Chapter 14) (Remy *et al.*, 1994). Weathering of soil enhanced the P availability for plants and microorganisms. During soil development (pedogenesis), however, soil P concentrations decline to much lower levels, due to long-term weathering, erosion and leaching (Turner & Condron, 2013; Walker & Syers, 1976). As a result, alternative mechanisms evolved to enhance the P-acquisition efficiency of plants on severely P-impooverished soils, including the root secretion of hydrolytic enzymes such as nucleases and acid phosphatases that mobilise inorganic-P (Pi) from the soil's organic-P pool (Chapter 10) (Plaxton & Tran, 2011), as well as the release of carboxylates that mobilise P from both organic-P and (Pi) sources (Chapter 11) (Lambers *et al.*, 2006; Lambers *et al.*, 2008).

Once P is liberated from minerals during weathering, it is quickly sequestered into a number of more recalcitrant phases (Walker & Syers, 1976), limiting its availability to plants and microorganisms (Chapter 13) (Lambers *et al.*, 2009; Porder *et al.*, 2007). Unlike the situation for nutrients such as nitrogen and carbon, ecosystems depend entirely on the aqueous transfer of P. Notable exceptions are P-impooverished ecosystems that rely on the import of Aeolian dust, for example the Amazon Basin (Bristow *et al.*, 2009; Bristow *et al.*, 2010) and old volcanic islands in Hawaii (Chadwick *et al.*, 1999).

Variations in the global P cycle occurred not only when the atmospheric oxygen concentration was increased (Canfield *et al.*, 2013), but also, for

example, during the uplift of the Himalayan-Tibetan Plateau which increased chemical weathering, leading to an enhanced input of P to the oceans. Additionally, P is redistributed on glacial timescales, resulting from loss of the continental margin during glacial low sea levels (Filippelli, 2008). Global phosphate rock reserves are the result of the formation of P-bearing minerals in marine sediments; this process occurs both in continental margin sediments and in deep sea sediments (oozes) (Filippelli, 2011). Current rock phosphate reserves have been formed over millions of years, and are gradually being depleted (Cordell *et al.*, 2009). As rock P reserves decline (Fixen & Johnston, 2012), the costs of P fertilizers will rise, and this is a significant issue for farmers in developing nations that have limited access to P fertilizers (Lynch, 2007; Scholz & Wellmer, 2013). Improvements in the efficiency of cropping systems to acquire and use P are therefore urgently needed (Johnston *et al.*, 2014; Rose *et al.*, 2011).

1.4 Phosphorus as an essential, but frequently limiting, soil nutrient for plant productivity

Soil P concentrations would have been low soon after terrestrial life on Earth began, as outlined above for soils following thousands of years of pedogenesis. In the current era, in young soils, for example following the retreat of glaciers (Richardson *et al.*, 2004) or the deposition of coastal dunes (Laliberté *et al.*, 2013), soil P concentrations are relatively high and P is not the key limiting nutrient for plant growth. However, during pedogenesis, P rapidly becomes the major macronutrient limiting plant productivity (Vitousek *et al.*, 2010).

Pierre and Parker (1927) measured an average P_i concentration in the soil solution of 3 μM in a study of 21 different soils from the South and Middle West of the USA (range: <0.6 to 11 μM), far lower than the intracellular P_i concentrations (5–20 mM) required for optimal crop growth (Fang *et al.*, 2009; Vance *et al.*, 2003). The soils were chosen to represent a wide range in texture and organic matter content, and the average organic-P concentration was considerably higher: 15 μM (Pierre & Parker, 1927). More recent publications, including those dealing with soils fertilised with P at rates that are common agronomic practice, confirmed the range of soil P concentrations reported in this early work (Hossner *et al.*, 1973; Johnston *et al.*, 2014; Ron Vaz *et al.*, 1993; Yanai, 1991).

The P_i in the soil solution to sustain near-maximum growth of pearl millet (*Pennisetum typhoides*) is 6.5 μM (Fox & Kamprath, 1970), very similar to what has been found for a range of other crop and pasture species grown in nutrient solution (Asher & Loneragan, 1967; Breeze *et al.*, 1984; Breeze *et al.*, 1985). Even lower concentrations were found to saturate the growth of perennial ryegrass (*Lolium perenne*) in pot trials using phosphated-goethite as the source of P (Parfitt, 1979).

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The concentrations in the soil solution and used in experimental glasshouse experiments are remarkably low compared with those commonly used in experiments on P nutrition of, for example, *Arabidopsis thaliana* and tomato (*Solanum lycopersicum*), in which 250 μM (Dong *et al.*, 1998; Muchhal *et al.*, 1996; Muchhal & Raghothama, 1999), 1 mM (Narang *et al.*, 2000), or even 2.5 mM (Wu *et al.*, 2003) is 'normal' and 0 mM is commonly used as 'low'. Similarly high P concentrations (300–320 μM or 1 mM) are frequently used for the experimental growth of rice (*Oryza sativa*) (Li *et al.*, 2010; Ni *et al.*, 1996; Secco *et al.*, 2013; Seo *et al.*, 2008) under 'normal' conditions, and 16 or 35 μM for growth at a 'low' P availability (Ni *et al.*, 1996; Seo *et al.*, 2008). However, it would be incorrect to assume that these high reported concentrations are really present as free P_i ions in solution (either in agar or in the liquid culture medium). It is impossible to achieve such high concentrations whenever similarly high calcium concentrations are used in the culture medium, because calcium phosphates precipitate. The real P_i concentration in solution, as opposed to the total amount of P, can be calculated using Geochem-PC (Parker *et al.*, 1995; Shaff *et al.*, 2010). In agar, the adsorption of P_i onto the gel, as well as the agar being a potential source of additional P at low P loadings, offer additional complications; Eurobio agarose may contain up to 43 $\mu\text{g P g}^{-1}$ dry weight of agarose (Irshad *et al.*, 2012) and other sources of agar even more (U. Irshad and C. Plassard, pers. comm.). With respect to the 'low-P' plants, in reality, these will have been exposed to a concentration supplied at the start (e.g. 16 or 35 μM in the cited examples for rice) and 'zero P' when they exhausted all P in the nutrient solution. It takes growing conditions with a very large volume or very frequent replacement of nutrient solutions to really maintain a low P concentration in solution for extended periods (Asher & Loneragan, 1967; Asher, 1981; Breeze *et al.*, 1984). An alternative approach is that developed by Ingestad (1970; 1982), who imposed an exponential growth rate on seedlings by increasing the relative addition rate of a limiting nutrient exponentially. In this way, plants can be grown in a steady state whilst being limited by a specific nutrient, such as P (De Groot *et al.*, 2001; Ericsson & Ingestad, 1988).

The 'normal' P_i concentrations used in nutrient solution, for example 0.1 mM P_i for the growth of barley (*Hordeum vulgare*) or barleygrass (*H. leporinum*), may not cause P-toxicity problems because the P_i -uptake systems in these species are downregulated, but they do lead to an excessive accumulation of P_i in leaves (Chapin & Bielecki, 1982). In species such as *Hakea prostrata*, that do not have a less-pronounced capacity to down-regulate P-uptake systems, there may well be a risk of P toxicity. In other species, such as *Trifolium subterraneum* (subterranean clover) and *Lupinus digitatus* (blue lupin) (Asher & Loneragan, 1967), a range of tropical food legumes (*Cajanus cajan* (pigeonpea), *Pachyrrhizus erosus* (Mexican yam bean), *Psophocarpus tetragonolobus* (winged bean), *Vigna angularis* (adzuki bean), *V. mungo* (black gram), *V. unguiculata* (cowpea) (Bell *et al.*, 1990) and *Medicago*

truncatula (Tang *et al.*, 2001), toxicity symptoms occur at 15 or 25 μM Pi in nutrient solution, especially – but not exclusively – when the nitrogen supply is suboptimal. Toxicity symptoms are often observed when plants, for example *Solanum tuberosum* (potato) (Cogliatti & Clarkson, 1983) or *H. vulgare* (Green *et al.*, 1973), are pregrown in a low-P solution and then exposed to a ‘normal’ P solution. If we seek to study plants with a realistic P status, it is essential to supply Pi in a manner that avoids both an accumulation of Pi to very high levels and a depletion of Pi in the nutrient solution. In addition to the approaches discussed in the preceding paragraph, plants can be grown in soil or sand, but even then care must be taken not to provide unrealistically high Pi concentrations.

1.5 Soil phosphorus pools

The Pi pool in the soil solution tends to be small (Pierre & Parker, 1927), but the total inorganic soil P pool varies over three orders of magnitude, mainly depending on soil age (Turner *et al.*, 2013). In ancient weathered soils in southwestern Australia, the readily available Pi concentration can be as low as 1 mg kg^{-1} or less (Lambers *et al.*, 2012), whereas in young volcanic soil in Chile, values of 1000 mg kg^{-1} are common (Borie & Rubio, 2003). The Pi pool in the soil solution is readily available for uptake by plants, but that is not the case for a large fraction of the Pi pool, which can be strongly sorbed to oxides and hydroxides of iron and aluminium (Barrow, 1999; Borie & Rubio, 2003). The extent to which the soil’s total Pi pool can be accessed by plants using different P-acquisition strategies will be explored in detail in several chapters of this book (see Chapters 10, 11, 13, 14).

Phosphorus is not distributed homogeneously through the soil profile, except in very young soils such as recently deposited sand dunes (Laliberté *et al.*, 2012). As soils develop and plants remove nutrients from them, plant litter is deposited on the soil surface and thus a soil P profile develops, with highest concentrations in the surface layers (Laliberté *et al.*, 2012; Smeck, 1973; Walbridge *et al.*, 1991). In agricultural soils, a similar profile is to be expected, as a result of manure or inorganic-P fertilizer being spread on top of the soil (Dick, 1983; Holanda *et al.*, 1998), especially under no-tillage conditions (Cade-Menun *et al.*, 2010; Guertal *et al.*, 1991). Weathering and pedogenesis result in a decrease of total P, especially Pi, with organic-P becoming relatively more important (Walker & Syers, 1976). This is most pronounced in the topsoil, which is the most weathered soil horizon in a soil profile, relative to deeper soil horizons (Turner *et al.*, 2013).

The organic-P concentration in the soil solution can be about fivefold greater than the Pi pool (Pierre & Parker, 1927). The total organic-P pool as a fraction of the total P concentration is very small (<0.1%) on very young soils along the Franz Josef chronosequence in New Zealand, but

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that fraction increases to about half of the total P concentration in older soils (Turner *et al.*, 2013). In lowland tropical rainforest soils, organic-P represents 26% of total soil P (Turner & Engelbrecht, 2011). whereas in high-P volcanic soils in Chile organic-P constitutes about half of the total soil P (Borie & Rubio, 2003). The organic-P pool comprises a range of chemical compounds: phosphate monoesters (e.g. inositol phosphates, sugar phosphates and mononucleotides) and phosphate diesters (e.g. phospholipids, DNA), organic-polyphosphates, phosphonates, and phytates (Cade-Menun *et al.*, 2010; Chapuis-Lardy *et al.*, 2001; Turner *et al.*, 2005; Turner & Engelbrecht, 2011).

Although plants take up Pi (as discussed in Chapter 5), they can also access a fraction of the organic-P pool, following hydrolysis of some organic-P compounds by secreted nucleases, phosphodiesterases, purple acid phosphatases (Chapter 10), and phytases (George *et al.*, 2006a; George *et al.*, 2006b; Maruyama *et al.*, 2012; Plaxton & Tran, 2011; Tarafdar & Claassen, 1988). These Pi-releasing enzymes may be excreted by the roots themselves, or they may be of microbial origin (Chapter 13) (Kitayama, 2013).

1.6 Soil phosphorus mobility

Because of the low concentrations of Pi in the soil solution and the high reactivity of Pi, minute amounts of Pi move via mass flow towards the root surface of transpiring plants. Mass flow typically delivers as little as 1–5% of a plant's P demand (Barber, 1962; Oliveira *et al.*, 2010; Prenzel, 1979), and the amount intercepted by growing roots is even less than that (Barber *et al.*, 1963; Clarkson, 1981). Organic-P concentrations in the soil solution tend to be higher than Pi concentrations (Pierre & Parker, 1927), which likely contributes to their greater mobility in soil (Hannapel *et al.*, 1964a). In addition, microbial activity – presumably the conversion of immobile organic-P into mobile organic-P – may account for greater mobility of organic-P, because the stimulation of microbial activity enhances the mobility of organic-P, whereas its inhibition decreases its mobility (Hannapel *et al.*, 1964b).

Most Pi arrives at the root surface by diffusion (Bhadoria *et al.*, 1991a; Bhat & Nye, 1973; Drew & Nye, 1970), followed by active transport across the plasma membrane of root hair and root epidermal cells (Chapter 5) or of mycorrhizal fungal hyphae (Chapter 14). However, the diffusion coefficient of Pi in soil is relatively low, compared to that of some other nutrients, typically of the order of $0.1 - 5 \times 10^{-13} \text{ m}^2 \text{ s}^{-1}$. Since this diffusion coefficient declines with decreasing soil moisture content (Bhadoria *et al.*, 1991b), any root activity that increases the moisture content in the rhizosphere will potentially increase plant P acquisition (as discussed in Section 1.7). Plants have a range of mechanisms, which enhance the acquisition of sufficient Pi to sustain their growth, many of which will be elaborated on in subsequent chapters (Chapters 11–14).

1.7 Factors determining rates of phosphorus uptake by roots

Given the very low mobility of Pi in dry soil, plant activities that enhance this mobility are expected to increase plant Pi uptake. The release of water by roots into superficial dry soil layers, taken up from moist deeper layers (hydraulic lift), is therefore expected to increase Pi acquisition (Prieto *et al.*, 2012). Likewise, the release of phospholipid surfactants by roots changes the biophysical properties of the rhizosphere and increases soil solution Pi concentration (Read *et al.*, 2003).

Whilst mass flow contributes very little to the acquisition of Pi by crop plants, it is possible that this situation is different for slow-growing plants in sandy soils (Cernusak *et al.*, 2011; Matimati *et al.*, 2014). However, such sandy soils contain very little Pi, so if mass flow is to have an effect on P acquisition, it is likely to involve organic-P forms, some of which are more mobile in soil (Frossard *et al.*, 1989; Hoffman & Rolston, 1980). Unless the concentration of organic-P in the soil solution is very high and the plant's P demand is relatively low, it is unlikely that mass flow can deliver a substantial component of the plant's P requirement. Root activities that enhance Pi in the soil solution can be expected to have a major impact on Pi uptake (see Chapters 11 and 13) (Lambers *et al.*, 2006).

Simulation models of plant P uptake that take into account both soil and root characteristics have been used to assess which traits have a major impact on net P uptake (Figure 1.2) (Schenk & Barber, 1979; Silberbush & Barber, 1983). The rate of root elongation and root diameter, including root hairs (Bhat & Nye, 1973), are among the most important root traits. What was not included in the early models was the nonhomogeneous distribution of P in the soil profile, but this aspect has been addressed recently (Dunbabin *et al.*, 2013). Jonathan Lynch and coworkers acknowledged the distribution of P in the soil profile, and suggested selecting for genotypes with shallow roots that would readily access poorly mobile nutrients in shallow soil layers (Lynch & Brown, 2008; Postma & Lynch, 2012; York *et al.*, 2013). What was also not included in the early simulation models is the role of mycorrhizal hyphae in P acquisition. When roots are colonised by arbuscular mycorrhizal fungi, the Pi transporters in their epidermal cells that are responsible for Pi uptake from the rhizosphere are downregulated, and Pi uptake by the mycorrhizal hyphae is the dominant pathway for P acquisition (Chapter 14) (Smith *et al.*, 2003). Transporters that are inducible by arbuscular mycorrhizal fungi are expressed in cortical cells, acquiring Pi released by arbuscules (Karandashov & Bucher, 2005; Paszkowski *et al.*, 2002). The mycorrhiza-inducible Pi transporter genes are downregulated at high Pi supply (Nagy *et al.*, 2009). Beyond mycorrhizal hyphae, P acquisition by plants largely relies on processes of mobilisation of poorly soluble forms of inorganic and organic-P in the rhizosphere – that is, on many biogeochemical and biochemical processes driven by roots or rhizosphere microbiota (Chapter 13) (Clarkson, 1985; Hinsinger

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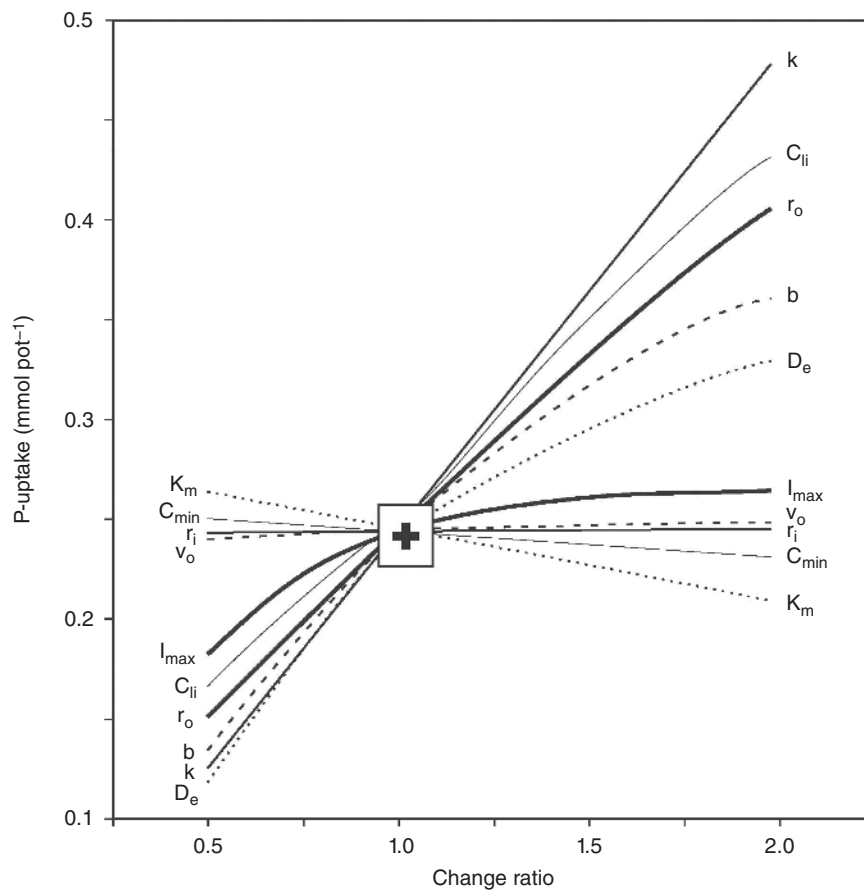


Figure 1.2 Effects of changing parameter values (from 0.5- to 2.0-fold the standard value) on simulated inorganic phosphate (Pi) uptake by roots of soybean (*Glycine max*). k is the rate of root elongation; C_{ii} is the initial Pi concentration in the soil solution; r_o is the root diameter; b is the buffer power of the soil; D_e is the diffusion coefficient of Pi in the soil; I_{max} is the maximum Pi inflow rate; v_o is the rate of transpiration; r_i is the spacing between individual roots; C_{min} is the lowest concentration at which Pi uptake is possible; and K_m is the Pi concentration at which the rate of Pi uptake is 50% of I_{max} (Silberbush & Barber, 1983).

et al., 2001). Most plant nutrition models do not account for these key processes, however, which explains why they strongly underestimate the actual P acquisition under limiting soil P conditions (Hinsinger *et al.*, 2011).

The least important traits for P acquisition from soil, according to the simulation model of Silberbush and Barber (1983), are the kinetic properties of the root's P-uptake system (Figure 1.2). This is to be expected, given that P mobility in soil is a major constraint for P uptake from soil (Clarkson, 1985; Tinker & Nye, 2000). Remarkably, this major result is not taken on board by many

who work on P-starvation responses. Of course, high-affinity Pi transporters are crucially important for Pi uptake from the rhizosphere. As discussed in Chapter 5, the enhanced expression of genes encoding high-affinity Pi transporters during P starvation is considered important to increase Pi uptake, whereas the results shown in Figure 1.2 indicate it is obviously not. How can this discrepancy be resolved? The actual significance of the change in expression of the genes encoding Pi transporters that are involved in Pi uptake from the rhizosphere is not their 'upregulation' at limiting P supply, but their 'downregulation' at very high Pi supply. Upregulations and downregulations are simply two ends of the same continuum. Species that occur naturally in P-impooverished environments with a very low capacity to downregulate their Pi-uptake system are very sensitive to P toxicity (de Campos *et al.*, 2013; Shane *et al.*, 2004). Interestingly, one such P-sensitive species, *Eucalyptus marginata* (jarrah), only shows severe P-toxicity symptoms following a toxic Pi pulse when it is not colonised by mycorrhizal fungi (Kariman *et al.*, 2014), which suggests that its Pi-uptake systems are downregulated only when the plants are mycorrhizal. Other Pi-sensitive plants include the *Pho2* mutant of *Arabidopsis thaliana* (Dong *et al.*, 1998), which is defective in a ubiquitin-conjugating E2 enzyme (Liu *et al.*, 2012) and a transgenic overexpressing miR399 (Aung *et al.*, 2006). These plants do not downregulate their Pi-uptake systems, and accumulate excessive amounts of Pi in their shoots (Chapter 2) (Lin *et al.*, 2008). In soils where the mobility of P is very high, such as rice paddy fields, the kinetic properties of the Pi-uptake system may be important (Park *et al.*, 2007; Seo *et al.*, 2008), but further studies are required to confirm this point.

1.8 Phosphorus-starvation responses: does phosphorus homeostasis exist?

Plants show a range of responses to a low P supply which are generally referred to as 'P-starvation responses' (e.g. Karthikeyan *et al.*, 2014; Plaxton & Tran, 2011; Ticconi *et al.*, 2001; Yang & Finnegan, 2010). These P-starvation responses minimise plant P deficiency, and include decreased growth and increased root/shoot ratio, root-hair density and carboxylate exudation (Lambers *et al.*, 2006). They also involve a decrease in the uptake and metabolism of nitrogen (Chapter 7) (e.g. Gniazdowska *et al.*, 1999; Rufty *et al.*, 1993). It is often claimed that these P-starvation responses lead to P homeostasis, but does that suggestion make sense? Before that question can be answered, it is important to confirm what 'homeostasis' really means. Based on the suggestions of Cannon (1929), who coined the term but gives full credit to Claude Bernard for the concept, Wikipedia captures it in an excellent manner: "From homoeostasis or homœostasis (from Greek: ὁμοιος, 'hómoios', 'similar', and στάσις, 'stásis', 'standing still') — is the property of a system in which variables are regulated so that inter-

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nal conditions remain stable and relatively constant. Examples of homeostasis include the regulation of temperature and the balance between acidity and alkalinity (pH). It is a process that maintains the stability of the human body's internal environment in response to changes in external conditions" (<http://en.wikipedia.org/wiki/Homeostasis>). Following this perfectly clear definition, there can be no P-starvation responses and homeostasis at the same time. When plants are grown at a limiting P supply, they dramatically decrease their tissue P concentration, especially the Pi concentration (Veneklaas *et al.*, 2012), and thus do not maintain homeostasis. Initially, there is a depletion of the vacuolar Pi pool, followed by a large (up to 50-fold) reduction in cytosolic Pi and organic-P levels (Veneklaas *et al.*, 2012). What the P-starvation response does, therefore, is to help stave off rapid cell death that would otherwise ensue, as for example reported for P-sufficient *Brassica napus* suspension cells cultured into media without Pi. The cells stop growing after a few days, but otherwise remain at least 90% viable for about three to four weeks. Blocking the P-starvation response by adding 1 mM phosphite to the medium causes them to enter programmed cell death within a few days, and the cells die within a week (Singh *et al.*, 2003). There is nothing wrong with either Claude Bernard's concept of homeostasis or with the concept of a P-starvation response, but the two terms are incompatible, and it is not helpful to use the term homeostasis when dealing with plant functioning in response to low-P conditions. If it were to be specified that homeostasis refers to the cytosolic Pi concentration, that would be a different matter, but that would need to be spelled out, which is commonly not done.

1.9 Concluding remarks

When it comes to plant P nutrition, which is pivotal for much what is discussed throughout this book, it is clear that there is not much P on planet Earth, it does not rapidly move in the soil and, as a result, soil P is not readily accessible for most plants. Modern agriculture relies heavily on mined phosphate rock to produce fertilizer, but this is obviously a nonrenewable resource that is being depleted (Cordell *et al.*, 2009; Vance *et al.*, 2003). Whilst it is unlikely that 'peak phosphorus' will be reached during the next few decades (Fixen & Johnston, 2012), this does not negate the need to work towards more P-efficient and sustainable food-production systems (Johnston *et al.*, 2014; Scholz & Wellmer, 2013) with less impact on natural ecosystems (Heckrath *et al.*, 1995; Smith & Schindler, 2009). Moreover, we only have to go back to when rock phosphate was first used to fertilise crops to significantly enhance crop yield. It was soon discovered that some sources of rock phosphate contained significant amounts of cadmium found naturally in the organic-rich marine sediments that are precursors to rock phosphate (Filippelli, 2002). Whilst these cadmium levels do not affect plant productivity, they are harmful for consumers of the products from these plants, and hence the

food chain (Chaney, 2012; Chaney, 2013; Grant & Sheppard, 2008). Having plenty of phosphate rock reserves does not mean that these resources are of similar quality as what is used today; some may contain high concentrations of heavy metals, or may be difficult and expensive to mine, and increasing fertilizer prices can therefore be anticipated (Scholz & Wellmer, 2013). Peak P therefore deserves a place on the political agenda, and represents a subject that plant and soil scientists need to bring to the attention of a wider audience.

The various chapters in this book explore plant P nutrition from various angles, enhancing our fundamental understanding of how plants acquire and use this essential nutrient (Chapters 2–4), and how they remobilise P from senescing tissues (Chapters 6 and 10). This will, in turn, allow the development of effective biotechnological strategies to produce plants that are more efficient at either acquiring or using P, and thus enhance the efficiency of cropping systems.

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References

- Andersson, H., Bergström, L., Djodjic, F. *et al.* (2013). Topsoil and subsoil properties influence phosphorus leaching from four agricultural soils. *Journal of Environmental Quality* **42**, 455–463.
- Asher, C.J. & Loneragan, J.F. (1967). Response of plants to phosphate concentration in solution culture: I. Growth and phosphorus content. *Soil Science* **103**, 225–233.
- Asher, C.J. (1981). Limiting external concentrations of trace elements for plant growth: use of flowing solution culture techniques. *Journal of Plant Nutrition* **3**, 163–180.
- Aung, K., Lin, S.-I., Wu, C.-C. *et al.* (2006). *pho2*, a phosphate overaccumulator, is caused by a nonsense mutation in a microRNA399 target gene. *Plant Physiology* **141**, 1000–1011.
- Barber, S.A. (1962). A diffusion and mass-flow concept of soil nutrient availability. *Soil Science* **93**, 39–49.
- Barber, S.A., Walker, J.M. & Vasey, E.H. (1963). Mechanisms for movement of plant nutrients from soil and fertilizer to plant root. *Journal of Agricultural and Food Chemistry* **11**, 204–207.
- Barrow, N.J. (1999). A simple equation to describe sorption of anions by goethite. *European Journal of Soil Science* **50**, 151–155.
- Bekker, A. & Holland, H.D. (2012). Oxygen overshoot and recovery during the early Paleoproterozoic. *Earth and Planetary Science Letters* **317–318**, 295–304.

16 ■ Phosphorus Metabolism in Plants

- Bell, R.W., Edwards, D.G. & Asher, C.J. (1990). Growth and nodulation of tropical food legumes in dilute solution culture. *Plant and Soil* **122**, 249–258.
- Bhadoria, P.B.S., Kaselowsky, J., Claassen, N. *et al.* (1991a). Soil phosphate diffusion coefficients: their dependence on phosphorus concentration and buffer power. *Soil Science Society of America Journal* **55**, 56–60.
- Bhadoria, P.B.S., Kaselowsky, J., Claassen, N. *et al.* (1991b). Phosphate diffusion coefficients in soil as affected by bulk density and water content. *Zeitschrift für Pflanzenernährung und Bodenkunde* **154**, 53–57.
- Bhat, K.K.S. & Nye, P.H. (1973). Diffusion of phosphate to plant roots in soil. *Plant and Soil* **38**, 161–175.
- Borie, F. & Rubio, R. (2003). Total and organic phosphorus in Chilean volcanic soils. *Gayana Botanica* **60**, 69–73.
- Breeze, V.G., Wild, A., Hopper, M.J. *et al.* (1984). The uptake of phosphate by plants from flowing nutrient solution: II. Growth of *Lolium perenne* L. at constant phosphate concentrations. *Journal of Experimental Botany* **35**, 1210–1221.
- Breeze, V.G., Robson, A.D. & Hopper, M.J. (1985). The uptake of phosphate by plants from flowing nutrient solution: III. Effects of changed phosphate concentrations on the growth and distribution of phosphate within plants of *Lolium perenne* L. *Journal of Experimental Botany* **36**, 725–733.
- Bristow, C.S., Drake, N. & Armitage, S. (2009). Deflation in the dustiest place on Earth: the Bodélé Depression, Chad. *Geomorphology* **105**, 50–58.
- Bristow, C.S., Hudson-Edwards, K.A. & Chappell, A. (2010). Fertilizing the Amazon and equatorial Atlantic with West African dust. *Geophysical Research Letters* **37**, L14807.
- Cade-Menun, B.J., Carter, M.R., James, D.C. *et al.* (2010). Phosphorus forms and chemistry in the soil profile under long-term conservation tillage: a phosphorus-31 nuclear magnetic resonance study. *Journal of Environment Quality* **39**, 1647–1656.
- Canfield, D.E., Ngombi-Pemba, L., Hammarlund, E.U. *et al.* (2013). Oxygen dynamics in the aftermath of the Great Oxidation of Earth's atmosphere. *Proceedings of the National Academy of Sciences of the United States of America* **110**, 16736–16741.
- Cannon, W.B. (1929). Organisation for physiological homeostasis. *Physiological Reviews* **9**, 399–422.
- Carswell, C., Grant, B.R., Theodorou, M.E. *et al.* (1996). The fungicide phosphonate disrupts the phosphate-starvation response in *Brassica nigra* seedlings. *Plant Physiology* **110**, 105–110.
- Cernusak, L.A., Winter, K. & Turner, B.L. (2011). Transpiration modulates phosphorus acquisition in tropical tree seedlings. *Tree Physiology* **31**, 878–885.
- Chadwick, O.A., Derry, L.A., Vitousek, P.M. *et al.* (1999). Changing sources of nutrients during four million years of ecosystem development. *Nature* **397**, 491–497.
- Chaney, R.L. (2012). Food safety issues for mineral and organic fertilizers. *Advances in Agronomy* **117**, 51–116.
- Chaney, R.L., Ryan, J.A. & Reeves, P.G. (2013). *Cadmium in soils and its transfer to plants and the human food chain*. Paper presented at the Cadmium 2001: Proceedings, Eighth International Cadmium Conference, Kunming, China.
- Chapin, F.S. & Bielecki, R.L. (1982). Mild phosphorus stress in barley and a related low-phosphorus-adapted barleygrass: phosphorus fractions and phosphate absorption in relation to growth. *Physiologia Plantarum* **54**, 309–317.

- Chapuis-Lardy, L., Brossard, M. & Quiquampoix, H. (2001). Assessing organic phosphorus status of Cerrado oxisols (Brazil) using ^{31}P -NMR spectroscopy and phosphomonoesterase activity measurement. *Canadian Journal of Soil Science* **81**, 591–601.
- Clarkson, D.T. (1981). Nutrient interception and transport by root systems. In: Johnson, C.B. (ed.) *Physiological Processes Limiting Plant Productivity*, Butterworths, London, pp. 307–314.
- Clarkson, D.T. (1985). Factors affecting mineral nutrient acquisition by plants. *Annual Review of Plant Physiology* **36**, 77–115.
- Cogliatti, D.H. & Clarkson, D.T. (1983). Physiological changes in, and phosphate uptake by potato plants during development of, and recovery from phosphate deficiency. *Physiologia Plantarum* **58**, 287–294.
- Cooper, J., Lombardi, R., Boardman, D. *et al.* (2011). The future distribution and production of global phosphate rock reserves. *Resources, Conservation and Recycling* **57**, 78–86.
- Cordell, D., Drangert, J.-O. & White, S. (2009). The story of phosphorus: global food security and food for thought. *Global Environmental Change* **19**, 292–305.
- de Campos, M.C.R., Pearse, S.J., Oliveira, R.S. *et al.* (2013). Down-regulation of net phosphorus uptake capacity is inversely related to leaf phosphorus resorption in four species from a phosphorus-impooverished environment. *Annals of Botany* **111**, 445–454.
- De Groot, C.C., Marcelis, L.F.M., Van Den Boogaard, R. *et al.* (2001). Growth and dry-mass partitioning in tomato as affected by phosphorus nutrition and light. *Plant, Cell & Environment* **24**, 1309–1317.
- Dick, W.A. (1983). Organic carbon, nitrogen, and phosphorus concentrations and pH in soil profiles as affected by tillage intensity. *Soil Science Society of America Journal* **47**, 102–107.
- Dong, B., Rengel, Z. & Delhaize, E. (1998). Uptake and translocation of phosphate by *pho2* mutant and wild-type seedlings of *Arabidopsis thaliana*. *Planta* **205**, 251–256.
- Drew, M.C. & Nye, P.H. (1970). The supply of nutrient ions by diffusion to plant roots in soil. *Plant and Soil* **33**, 545–563.
- Dunbabin, V., Postma, J., Schnepf, A. *et al.* (2013). Modelling root–soil interactions using three-dimensional models of root growth, architecture and function. *Plant and Soil* **372**, 93–124.
- Dunne, C.P., Crane, C.E. & Shearer, B.L. (2010) *Stem application of phosphite controls Phytophthora cinnamomi in native plant communities from south-west Western Australia*. Paper presented at the Fifth Meeting of the International Union of Forest Research Organisations (IUFRO) Working Party, Rotorua, New Zealand, 7–12 March 2010.
- Ericsson, T. & Ingestad, T. (1988). Nutrition and growth of birch seedlings at varied relative phosphorus addition rates. *Physiologia Plantarum* **72**, 227–235.
- Fang, Z., Shao, C., Meng, Y. *et al.* (2009). Phosphate signaling in *Arabidopsis* and *Oryza sativa*. *Plant Science* **176**, 170–180.
- Filippelli, G.M. (2002). The global phosphorus cycle. *Reviews in Mineralogy and Geochemistry* **48**, 391–425.
- Filippelli, G.M. (2008). The global phosphorus cycle: past, present, and future. *Elements* **4**, 89–95.
- Filippelli, G.M. (2011). Phosphate rock formation and marine phosphorus geochemistry: the deep time perspective. *Chemosphere* **84**, 759–766.
- Fixen, P.E. & Johnston, A.M. (2012). World fertilizer nutrient reserves: a view to the future. *Journal of the Science of Food and Agriculture* **92**, 1001–1005.

18 ■ Phosphorus Metabolism in Plants

- Fox, R.L. & Kamprath, E.J. (1970). Phosphate sorption isotherms for evaluating the phosphate requirements of soils. *Soil Science Society of America Journal* **34**, 902–907.
- Frossard, E., Stewart, J.W.B. & Arnaud, R.J.S. (1989). Distribution and mobility of phosphorus in grassland and forest soils of Saskatchewan. *Canadian Journal of Soil Science* **69**, 401–416.
- George, T.S., Quiquampoix, H., Simpson, R.J. *et al.* (2006a). Interactions between phytases and soil constituents: implications for the hydrolysis of inositol phosphates. In: Turner, B.L. & Richardson, A.E. (eds) *Inositol Phosphates: Linking Agriculture and the Environment*. CAB International, pp. 222–242.
- George, T.S., Turner, B.L., Gregory, P.J. *et al.* (2006b). Depletion of organic phosphorus from Oxisols in relation to phosphatase activities in the rhizosphere. *European Journal of Soil Science* **57**, 47–57.
- Gniazdowska, A., Krawczak, A., Mikulska, M. *et al.* (1999). Low phosphate nutrition alters bean plants' ability to assimilate and translocate nitrate. *Journal of Plant Nutrition* **22**, 551–563.
- Grant, C.A. & Sheppard, S.C. (2008). Fertilizer impacts on cadmium availability in agricultural soils and crops. *Human and Ecological Risk Assessment: An International Journal* **14**, 210–228.
- Green, D.G., Ferguson, W.S. & Warder, F.G. (1973). Accumulation of toxic levels of phosphorus in the leaves of phosphorus-deficient barley. *Canadian Journal of Plant Science* **53**, 241–246.
- Greenwood, J.P. & Blake, R.E. (2006). Evidence for an acidic ocean on Mars from phosphorus geochemistry of Martian soils and rocks. *Geology* **34**, 953–956.
- Griffith, E.J., Ponnampereuma, C. & Gabel, N. (1977). Phosphorus, a key to life on the primitive earth. *Origins of Life* **8**, 71–85.
- Guertal, E.A., Eckert, D.J., Traina, S.J. *et al.* (1991). Differential phosphorus retention in soil profiles under no-till crop production. *Soil Science Society of America Journal* **55**, 410–413.
- Hannapel, R.J., Fuller, W.H., Bosma, S. *et al.* (1964a). Phosphorus movement in a calcareous soil: I. Predominance of organic forms of phosphorus in phosphorus movement. *Soil Science* **97**, 350–357.
- Hannapel, R.J., Fuller, W.H. & Fox, R.H. (1964b). Phosphorus movement in a calcareous soil: II. Soil microbial activity and organic phosphorus movement. *Soil Science* **97**, 421–427.
- Heckrath, G., Brookes, P.C., Poulton, P.R. *et al.* (1995). Phosphorus leaching from soils containing different phosphorus concentrations in the Broadbalk experiment. *Journal of Environmental Quality* **24**, 904–910.
- Hinsinger, P., Fernandes Barros, O.N., Benedetti, M.F. *et al.* (2001). Plant-induced weathering of a basaltic rock: experimental evidence. *Geochimica et Cosmochimica Acta* **65**, 137–152.
- Hinsinger, P., Brauman, A., Devau, N. *et al.* (2011). Acquisition of phosphorus and other poorly mobile nutrients by roots. Where do plant nutrition models fail? *Plant and Soil* **348**, 29–61
- Hoffman, D.L. & Rolston, D.E. (1980). Transport of organic phosphate in soil as affected by soil type. *Soil Science Society of America Journal* **44**, 46–52.
- Holanda, F.S.R., Mengel, D.B., Paula, M.B. *et al.* (1998). Influence of crop rotations and tillage systems on phosphorus and potassium stratification and root distribution in the soil profile. *Communications in Soil Science and Plant Analysis* **29**, 2383–2394.

- Hossner, L.R., Freeouf, J.A. & Folsom, B.L. (1973). Solution phosphorus concentration and growth of rice (*Oryza sativa* L.) in flooded soils. *Soil Science Society of America Journal* **37**, 405–408.
- Ingestad, T. (1970). A definition of optimum nutrient requirements in birch seedlings. I. *Physiologia Plantarum* **23**, 1127–1138.
- Ingestad, T. (1982). Relative addition rate and external concentration; driving variables used in plant nutrition research. *Plant, Cell and Environment* **5**, 443–453.
- Irshad, U., Brauman, A., Villenave, C. *et al.* (2012). Phosphorus acquisition from phytate depends on efficient bacterial grazing, irrespective of the mycorrhizal status of *Pinus pinaster*. *Plant and Soil* **358**, 155–168.
- Johnston, A.E., Poulton, P.R., Fixen, P.E. *et al.* (2014). Phosphorus: its efficient use in agriculture. *Advances in Agronomy* **123**, 177–228.
- Karandashov, V. & Bucher, M. (2005). Symbiotic phosphate transport in arbuscular mycorrhizas. *Trends in Plant Science* **10**, 22–29.
- Kariman, K., Barker, S.J., Finnegan, P.M. *et al.* (2014). Ecto- and arbuscular mycorrhizal symbiosis can induce tolerance to toxic pulses of phosphorus in jarrah (*Eucalyptus marginata*) seedlings. *Mycorrhiza* **24**, 501–509.
- Karthikeyan, A.S., Jain, A., Nagarajan, V.K. *et al.* (2014). *Arabidopsis thaliana* mutant lpsi reveals impairment in the root responses to local phosphate availability. *Plant Physiology and Biochemistry* **77**, 60–72.
- Kitayama, K. (2013). The activities of soil and root acid phosphatase in the nine tropical rain forests that differ in phosphorus availability on Mount Kinabalu, Borneo. *Plant and Soil* **367**, 215–224.
- Laliberté, E., Turner, B.L., Costes, T. *et al.* (2012). Experimental assessment of nutrient limitation along a 2-million year dune chronosequence in the south-western Australia biodiversity hotspot. *Journal of Ecology* **100**, 631–642.
- Laliberté, E., Turner, B.L., Zemunik, G. *et al.* (2013). Nutrient limitation along the Jurien Bay dune chronosequence: response to Uren & Parsons. *Journal of Ecology* **101**, 1088–1092.
- Lambers, H., Shane, M.W., Cramer, M.D. *et al.* (2006). Root structure and functioning for efficient acquisition of phosphorus: matching morphological and physiological traits. *Annals of Botany* **98**, 693–713.
- Lambers, H., Raven, J.A., Shaver, G.R. *et al.* (2008). Plant nutrient-acquisition strategies change with soil age. *Trends in Ecology and Evolution* **23**, 95–103.
- Lambers, H., Mougél, C., Jaillard, B. *et al.* (2009). Plant–microbe–soil interactions in the rhizosphere: an evolutionary perspective. *Plant and Soil* **321**, 83–115.
- Lambers, H., Cawthray, G.R., Giavalisco, P. *et al.* (2012). Proteaceae from severely phosphorus-impooverished soils extensively replace phospholipids with galactolipids and sulfolipids during leaf development to achieve a high photosynthetic phosphorus-use efficiency. *New Phytologist* **196**, 1098–1108.
- Li, L., Liu, C. & Lian, X. (2010). Gene expression profiles in rice roots under low phosphorus stress. *Plant Molecular Biology* **72**, 423–432.
- Lin, S.-I., Chiang, S.-F., Lin, W.-Y. *et al.* (2008). Regulatory network of microRNA399 and *PHO2* by systemic signaling. *Plant Physiology* **147**, 732–746.
- Liu, T.-Y., Huang, T.-K., Tseng, C.-Y. *et al.* (2012). *PHO2*-dependent degradation of *PHO1* modulates phosphate homeostasis in *Arabidopsis*. *Plant Cell* **24**, 2168–2183.
- Lynch, J.P. (2007). Roots of the second green revolution. *Australian Journal of Botany* **55**, 493–512.

20 ■ Phosphorus Metabolism in Plants

- Lynch, J.P. & Brown, K.M. (2008). Root strategies for phosphorus acquisition. In: *The Ecophysiology of Plant-Phosphorus Interactions*. Springer, pp. 83–116.
- Maruyama, H., Yamamura, T., Kaneko, Y. *et al.* (2012). Effect of exogenous phosphatase and phytase activities on organic phosphate mobilization in soils with different phosphate adsorption capacities. *Soil Science and Plant Nutrition* **58**, 41–51.
- Matimati, I., Verboom, G.A. & Cramer, M.D. (2014). Nitrogen regulation of transpiration controls mass-flow acquisition of nutrients. *Journal of Experimental Botany* **65**, 159–168.
- McDonald, A.E., Grant, B.R. & Plaxton, W.C. (2001). Phosphite (phosphorous acid): its relevance in the environment and agriculture and influence on plant phosphate starvation response. *Journal of Plant Nutrition* **24**, 1505–1519.
- Muchhal, U.S., Pardo, J.M. & Raghothama, K.G. (1996). Phosphate transporters from the higher plant *Arabidopsis thaliana*. *Proceedings of the National Academy of Sciences of the United States of America* **93**, 10519–10523.
- Muchhal, U.S. & Raghothama, K.G. (1999). Transcriptional regulation of plant phosphate transporters. *Proceedings of the National Academy of Sciences of the United States of America* **96**, 5868–5872.
- Nagy, R., Drissner, D., Amrhein, N. *et al.* (2009). Mycorrhizal phosphate uptake pathway in tomato is phosphorus-repressible and transcriptionally regulated. *New Phytologist* **181**, 950–959.
- Narang, R.A., Bruene, A. & Altmann, T. (2000). Analysis of phosphate acquisition efficiency in different *Arabidopsis* accessions. *Plant Physiology* **124**, 1786–1799.
- Ni, J.J., Wu, P., Lou, A.C. *et al.* (1996). Low phosphorus effects on the metabolism of rice seedlings. *Communications in Soil Science and Plant Analysis* **27**, 3073–3084.
- Oliveira, E.M.M., Ruiz, H.A., Alvarez, V.H. *et al.* (2010). Nutrient supply by mass flow and diffusion to maize plants in response to soil aggregate size and water potential. *Revista Brasileira de Ciência do Solo* **34**, 317–328.
- Parfitt, R.L. (1979). The availability of P from phosphate-goethite bridging complexes. Desorption and uptake by ryegrass. *Plant and Soil* **53**, 55–65.
- Park, M.R., Baek, S.-H., Reyes, B.G. *et al.* (2007). Overexpression of a high-affinity phosphate transporter gene from tobacco (*NtPT1*) enhances phosphate uptake and accumulation in transgenic rice plants. *Plant and Soil* **292**, 259–269.
- Parker, D.R., Norvell, W.A. & Chaney, R. (1995). GEOCHEM-PC—a chemical speciation program for IBM and compatible personal computers. *Chemical Equilibrium and Reaction Models* **42**, 253–269.
- Paszkowski, U., Kroken, S., Roux, C. *et al.* (2002). Rice phosphate transporters include an evolutionarily divergent gene specifically activated in arbuscular mycorrhizal symbiosis. *Proceedings of the National Academy of Sciences of the United States of America* **99**, 13324.
- Pierre, W.H. & Parker, F.W. (1927). Soil phosphorus studies: II. The concentration of organic and inorganic phosphorus in the soil solution and soil extracts and the availability of the organic phosphorus to plants. *Soil Science* **24**, 119–128.
- Plaxton, W.C. & Tran, H.T. (2011). Metabolic adaptations of phosphate-starved plants. *Plant Physiology* **156**, 1006–1015.
- Porder, S., Vitousek, P., Chadwick, O. *et al.* (2007). Uplift, erosion, and phosphorus limitation in terrestrial ecosystems. *Ecosystems* **10**, 159–171.
- Postma, J.A. & Lynch, J.P. (2012). Complementarity in root architecture for nutrient uptake in ancient maize/bean and maize/bean/squash polycultures. *Annals of Botany* **110**, 521–534.

- Prenzel, J. (1979). Mass flow to the root system and mineral uptake of a beech stand calculated from 3-year field data. *Plant and Soil* **51**, 39–49.
- Prieto, I., Armas, C. & Pugnaire, F.I. (2012). Water release through plant roots: new insights into its consequences at the plant and ecosystem level. *New Phytologist* **193**, 830–841.
- Ratjen, A.M. & Gerendás, J. (2009). A critical assessment of the suitability of phosphite as a source of phosphorus. *Journal of Plant Nutrition and Soil Science* **172**, 821–828.
- Read, D.B., Bengough, A.G., Gregory, P.J. *et al.* (2003). Plant roots release phospholipid surfactants that modify the physical and chemical properties of soil. *New Phytologist* **157**, 315–326.
- Remy, W., Taylor, T.N., Hass, H. *et al.* (1994). Four hundred-million-year-old vesicular arbuscular mycorrhizae. *Proceedings of the National Academy of Sciences of the United States of America* **91**, 11841.
- Richardson, S., Peltzer, D., Allen, R. *et al.* (2004). Rapid development of phosphorus limitation in temperate rainforest along the Franz Josef soil chronosequence. *Oecologia* **139**, 267–276.
- Ron Vaz, M.D., Edwards, A.C., Shand, C.A. *et al.* (1993). Phosphorus fractions in soil solution: influence of soil acidity and fertiliser additions. *Plant and Soil* **148**, 175–183.
- Rose, T.J., Rose, M.T., Pariasca-Tanaka, J. *et al.* (2011). The frustration with utilization: why have improvements in internal phosphorus utilization efficiency in crops remained so elusive? *Frontiers in Plant Nutrition* **2**, doi: 10.3389/fpls.2011.00073.
- Rufty, T.W., Israel, D.W., Volk, R.J. *et al.* (1993). Phosphate regulation of nitrate assimilation in soybean. *Journal of Experimental Botany* **44**, 879–891.
- Schenk, M.K. & Barber, S.A. (1979). Phosphate uptake by corn as affected by soil characteristics and root morphology. *Soil Science Society of America Journal* **43**, 880–883.
- Scholz, R.W. & Wellmer, F.-W. (2013). Approaching a dynamic view on the availability of mineral resources: what we may learn from the case of phosphorus? *Global Environmental Change* **23**, 11–27.
- Secco, D., Jabnourne, M., Walker, H. *et al.* (2013). Spatio-temporal transcript profiling of rice roots and shoots in response to phosphate starvation and recovery. *Plant Cell* **25**, 4285–4304.
- Seo, H.-M., Jung, Y., Song, S. *et al.* (2008). Increased expression of OsPT1, a high-affinity phosphate transporter, enhances phosphate acquisition in rice. *Biotechnology Letters* **30**, 1833–1838.
- Shaff, J.E., Schultz, B.A., Craft, E.J. *et al.* (2010). GEOCHEM-EZ: a chemical speciation program with greater power and flexibility. *Plant and Soil* **330**, 207–214.
- Shane, M.W., Szota, C. & Lambers, H. (2004). A root trait accounting for the extreme phosphorus sensitivity of *Hakea prostrata* (Proteaceae). *Plant, Cell and Environment* **27**, 991–1004.
- Silberbush, M. & Barber, S.A. (1983). Sensitivity of simulated phosphorus uptake to parameters used by a mechanistic-mathematical model. *Plant and Soil* **74**, 93–100.
- Simpson, R.J., Oberson, A., Culvenor, R.A. *et al.* (2011). Strategies and agronomic interventions to improve the phosphorus-use efficiency of farming systems. *Plant and Soil* **349**, 89–120.
- Singh, V., Wood, S., Knowles, V. *et al.* (2003). Phosphite accelerates programmed cell death in phosphate-starved oilseed rape (*Brassica napus*) suspension cell cultures. *Planta* **218**, 233–239.
- Smeck, N.E. (1973). Phosphorus: an indicator of pedogenetic weathering processes. *Soil Science* **115**, 199–206.

22 ■ Phosphorus Metabolism in Plants

- Smith, S.E., Smith, F.A. & Jakobsen, I. (2003). Mycorrhizal fungi can dominate phosphate supply to plants irrespective of growth responses. *Plant Physiology* **133**, 16–20.
- Smith, V.H. & Schindler, D.W. (2009). Eutrophication science: where do we go from here? *Trends in Ecology & Evolution* **24**, 201–207.
- Tang, C., Hinsinger, P., Drevon, J.J. *et al.* (2001). Phosphorus deficiency impairs early nodule functioning and enhances proton release in roots of *Medicago truncatula* L. *Annals of Botany* **88**, 131–138.
- Tarafdar, J. & Claassen, N. (1988). Organic phosphorus compounds as a phosphorus source for higher plants through the activity of phosphatases produced by plant roots and microorganisms. *Biology and Fertility of Soils* **5**, 308–312.
- Thao, H.T.B. & Yamakawa, T. (2009). Phosphite (phosphorous acid): fungicide, fertilizer or bio-stimulator? *Soil Science & Plant Nutrition* **55**, 228–234.
- Ticconi, C.A., Delatorre, C.A. & Abel, S. (2001). Attenuation of phosphate starvation responses by phosphite in *Arabidopsis*. *Plant Physiology* **127**, 963–972.
- Tinker, P.B. & Nye, P.H. (2000) *Solute Movement in the Rhizosphere*. Oxford University Press, New York, USA.
- Turner, B.L., Cade-Menun, B.J., Condrón, L.M. *et al.* (2005). Extraction of soil organic phosphorus. *Talanta* **66**, 294–306.
- Turner, B.L. & Engelbrecht, B. (2011). Soil organic phosphorus in lowland tropical rain forests. *Biogeochemistry* **103**, 297–315.
- Turner, B.L. & Condrón, L.M. (2013). Pedogenesis, nutrient dynamics, and ecosystem development: the legacy of T.W. Walker and J.K. Syers. *Plant and Soil* **367**, 1–10.
- Turner, B.L., Lambers, H., Condrón, L.M. *et al.* (2013). Soil microbial biomass and the fate of phosphorus during long-term ecosystem development. *Plant and Soil* **367**, 225–234.
- Vance, C.P., Uhde-Stone, C. & Allan, D.L. (2003). Phosphorus acquisition and use: critical adaptations by plants for securing a nonrenewable resource. *New Phytologist* **157**, 423–447.
- Veneklaas, E.J., Lambers, H., Bragg, J. *et al.* (2012). Opportunities for improving phosphorus-use efficiency in crop plants. *New Phytologist* **195**, 306–320.
- Vitousek, P.M., Porder, S., Houlton, B.Z. *et al.* (2010). Terrestrial phosphorus limitation: mechanisms, implications, and nitrogen-phosphorus interactions. *Ecological Applications* **20**, 5–15.
- Walbridge, M.R., Richardson, C.J. & Swank, W.T. (1991). Vertical distribution of biological and geochemical phosphorus subcycles in two southern Appalachian forest soils. *Biogeochemistry* **13**, 61–85.
- Walker, T.W. & Syers, J.K. (1976). The fate of phosphorus during pedogenesis. *Geoderma* **15**, 1–9.
- Wu, P., Ma, L., Hou, X. *et al.* (2003). Phosphate starvation triggers distinct alterations of genome expression in *Arabidopsis* roots and leaves. *Plant Physiology* **132**, 1260–1271.
- Yanai, R.D. (1991). Soil solution phosphorus dynamics in a whole-tree-harvested northern hardwood forest. *Soil Science Society of America Journal* **55**, 1746–1752.
- Yang, X.J. & Finnegan, P.M. (2010). Regulation of phosphate starvation responses in higher plants. *Annals of Botany* **105**, 513–526.
- York, L.M., Nord, E. & Lynch, J. (2013). Integration of root phenes for soil resource acquisition. *Frontiers in Plant Science* **4**.